CRISPR and Wildlife Conservation: Gene Editing as a Tool for Preserving Biodiversity

Debojani Narzary¹, Prof. Indira Boro²

¹Department of Zoology, University of Delhi, India ²Department of Bodo, Bodoland University, Assam, India

ABSTRACT

Genetic diversity is fundamental to the evolutionary potential of species. Due to the accelerating loss of biodiversity, there is an increased risk of species extinction; moreover, the traditional conservation methods are inadequate to safeguard vulnerable species. Therefore, conservation biology encourages emerging gene-editing tools, the CRISPR-Cas systems, which offer both opportunities and challenges. This review evaluates the application of CRISPR in wildlife conservation and ecosystem stability, while also highlighting the ethical, ecological and regulatory dimensions of gene editing in wild populations. Through case studies, we explore how CRISPR can be parallel to or rather surpass traditional methods in restoring genetic diversity. Ultimately, this review assesses both the potential and complexity of integrating genome editing tools with conservation frameworks responsibly, without underestimating the innovative use of CRISPR for the protection of biodiversity.

Keywords: Biodiversity, CRISPR-Cas9, conservation, ecosystem, genome-editing

INTRODUCTION

Rapid environmental changes caused by human impact have ushered in a novel crisis for global biodiversity. The accelerating rates of ecosystem degradation and species extinction are occurring up to 1,000 times faster than natural background levels (Pimm et al., 2014). According to reports of the IUCN Red List (2023), 42,100 species are threatened with extinction, representing more than 28% of all assessed species globally [1]. It reflects the urgency for global conservation strategies across all taxa and habitats. Some traditional methods such as captive breeding, habitat corridors and translocation are limited by long generation times, restricted gene pools, high operational costs, and difficulties in restoring lost alleles [2].

The CRISPR-Cas9 system is a revolutionary genome-editing tool that enables precise modification of DNA sequences in virtually any organism. Its adaptability, low cost and ease of use have made it a promising tool for wildlife conservation and ecosystem restoration. The system provides strategies to correct deleterious mutations, restore lost genetic variation, enhance disease resistance and control invasive species via synthetic gene drives. Although these strategies largely remain in a theoretical or experimental stage, they represent a significant shift in the field of conservation genetics [3]. This review explores the potential role of the CRISPR-Cas9 system in conserving wildlife by focusing on its applications in preserving the genetic diversity, while also addressing the ethical and ecological considerations required for its responsible use.

CRISPR-Cas SYSTEM

The CRISPR-Cas system - short for Clustered Regularly Interspaced Short Palindromic Repeats and CRISPR-associated proteins, originated as an adaptive immune mechanism in prokaryotes, where it protects against invading pathogens. CRISPR-Cas9 is one such system that is widely used for genome engineering across species. CRISPR-Cas9 consists of two components- a guide RNA (gRNA) which binds to target DNA and Cas9 endonuclease acting as a molecular scissor. The Cas9 protein when once it binds to the target site with gRNA, it induces a double-strand break in the DNA. The repair machinery of the cell then tries to repair the break via- non-homologous end joining (NHEJ) which often leads to insertions or deletions and homologous-directed repair (HDR) which allows gene insertion or correction precisely using a DNA template. The ease of its implementation along with precision has made CRISPR-Cas9 more potent than any other genome-editing tools like zinc finger nucleases (ZFNs) or transcription activator-like effector nucleases (TALENs) [4].

APPLICATIONS

Integration of CRISPR-Cas9 in wildlife conservation provides a novel method to precisely and effectively manipulate the genetic makeup ensuring adaptability, survivability of species and hence enhance ecosystem stability. Traditional methods are often lengthy, complex and more vulnerable to external factors.

A. Gene rescue in Endangered species

Endangered species are often faced with less genetic diversity due to historical bottlenecks, habitat fragmentation and inbreeding. These genetic issues then lead to deleterious alleles, reduced fecundity and developmental abnormalities [5]. Use of CRISPR technology can help repair these harmful alleles. For example, Black-footed ferret (Mustela Nigripes) is almost at the verge of extinction and efforts are being made to restore lost genetic variation using frozen cell lines from a now-extinct ferret named "Willa" whose genome was found to have unique alleles than the current population [6]. Theoretically, CRISPR can be used to knock-in these lost alleles and broaden the gene pool. Also, allele-specific correction with base editing and prime editing can minimize off-target effects- an advantage particularly for endangered species with restricted embryos for experimentation [7].

B. Enhancing disease resistance

Infectious diseases increase the risk of extinction of vulnerable species. For instance, Amphibians have seen a significant decline in their population due to chytridiomycosis, caused by the fungal pathogen Batrachochytrium dendrobatidis. Bat populations, similarly, are faced with white-nose syndrome in North America. Quarantines or antifungal treatments are often limited in wild settings.

CRISPR can be used to modify major histocompatibility complex (MHC) loci or antimicrobial peptide genes to confer resistance. A relevant example in agricultural species- pigs were given resistance to Porcine Reproductive and Respiratory Syndrome Virus (PRRSV) by deletion of a single host receptor gene [8]. This shows the potential of using CRISPR in protecting wild species.

C. Synthetic gene drives for Invasive species control

Small habitats such as islands where native species have not evolved resistance are majorly affected by invasive species. Traditional control strategies like trapping, poisoning, etc., are costly, labour-intensive and pose risks to non-target organisms.

By modifying traits (e.g., infertility, sex-ratio distortion), gene drives can spread deleterious traits among wild populations to eliminate the invasive species. For instance, this concept has been demonstrated in mosquitoes (Anopheles gambiae), where such gene drives that targeted female fertility achieved near-total population suppression in a caged environment. Modeling work suggests that gene drive strategies could be species-specific, self-propagating, and cost-effective [9]. However, real-world implementation is limited by gene flow to non-target populations, evolutionary resistance, and ecological unpredictability [10].

D. CRISPR for enhancing climate resilience: The case of Coral reefs

The mass bleaching events caused by the breakdown of the symbiosis between coral hosts and their dinoflagellate symbionts (Symbiodinium spp.), along with ocean warming and acidification, make the coral reefs most vulnerable among the ecosystems.

CRISPR can be used to enhance coral thermotolerance either by editing coral genomes or the symbionts' genomes. Levin et al.demonstrated CRISPR-mediated gene editing in reef-building coral Acropora millepora, where developmental and stress-response genes were targeted [11]. Genome-modified symbionts with improved thermal tolerance have been cultured and proposed for assisted evolution strategies [12].

E. De-extinction and the Revival of extinct phenotypes

De-extinction refers to the reconstruction of extinct phenotypes with the use of genome editing. The goal is not to recreate an identical organism but to reintroduce traits that were lost with extinction [13]. For instance, the Woolly Mammoth Revival project aims to introduce cold-adapted traits like subcutaneous fat, hair density, etc., into Asian elephants using CRISPR-Cas9[14]. Though it's controversial, the research has led to valuable studies on geneenvironment interactions.

In the context of conservation studies, partial de-extinction could allow rebuilding of degraded ecosystems with the introduction of species with traits like soil aeration, seed dispersal, browsing etc., required to stabilize the ecosystem.

F. Functional genomics of Non-model wildlife

Many non-model wildlife lacks a fully annotated genome which limits the understanding of trait heritability, adaptive pathways, and population dynamics. CRISPR enables the study of gene function in wild species with a reverse-genetics tool, further enabling conservation trait mapping. For example, CRISPR has been used to study coloration, beak morphology, and salinity tolerance in birds and fish. CRISPR enables the study of gene function in wild species with a reverse-genetics tool, further enabling conservation trait mapping and comparative genomics.

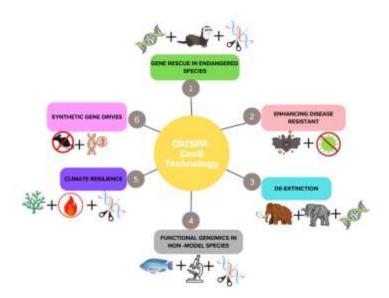


Figure 1: Conceptual overview of key applications of CRISPR-Cas9 technology in wildlife conservation.

RESEARCH GAPS

It becomes crucial to chart a strategic research plan to address scientific uncertainties, technological limitations, and sociopolitical complexities for responsible and effective implementation, as the application of the CRISPR-Cas system shifts from conceptual work to experimental models.

A. Integrating CRISPR with Conservation Genomics and Ecology

CRISPR should be viewed as part of an integrated conservation toolkit rather than in isolation. Combining gene editing with genomic selection, landscape genomics, and population viability analysis (PVA) can help identify genetically-informed management strategies.

Multidisciplinary research should focus on:

- Quantification of fitness consequences of specific modification in the wild
- Modeling gene flow and evolutionary trajectories post-editing.
- Study of ecological interactions across all trophic levels.

B. Stakeholder engagement and Ethical co-design

A critical need of governance models that incorporate the voices of indigenous groups, conservation NGOs, ecologists, ethicists, and the general public.

Social science research should focus on:

- Public attitudes towards genetic interventions in wild
- Cross-cultural values regarding species preservation and ecosystem integrity
- Mechanisms to achieve transparent, participatory decision-making

Initiatives such as the GeneConvene Global Collaborative seem to bridge these gaps, providing policy frameworks and resources for ethical gene drive deployment [15].

C. Establishing Long-Term Monitoring Protocols

There remains large speculation on the long-term impacts of CRISPR-based interventions on genetic diversity, ecosystem stability, and evolutionary dynamics.

Conservation biology must prioritize:

- Bio-surveillance systems development for edited wild populations.
- Create reversal protocols or "fail-safe" systems in gene drives.
- Long-term studies tracking multi-generational effects in field conditions.

D. Ethical Considerations in the use of the CRISPR-Cas system

As the margin between laboratory experiments and intervention in the wild blurs, the consequences of using CRISPR in wildlife conservation become increasingly complex. Genome edits are heritable and have self-propagating potential,

raising questions about ecological unpredictability and irreversibility. It can change the trajectories of evolution if such edited organisms are accidentally released [16]. The idea of "playing god" by interfering with natural evolutionary processes possesses a broader philosophical concern about the rights of humans to manipulate non-human species, even if the goals are conservation-oriented. Moreover, there is a risk of anthropocentric biases, valuing certain species or ecosystems over others based on aesthetics, tourism and political convenience.

CONCLUSION

The CRISPR-Cas system has revolutionizing potential to transform the direction of wildlife conservation with its precise, rapid, and cost-effective genomic interventions. From rescuing endangered populations to controlling invasive species, the applications are diverse and promising. Although it is required to use such a tool cautiously. Conservation requires collective efforts through interdisciplinary integration, ethical planning and public participation, and is not viewed solely as a biological endeavour. Both the methods of conservation, i.e., traditional methods and modern genomic technology, are mutually inclusive and they should be synergized for better flexibility and evidence-based measures. The proper exploitation of CRISPR in conservation depends on inclusive governance, long-term ecological surveillance, as well as just benefit-sharing frameworks. If responsibly implemented, CRISPR has the potential to change the direction of conservation strategies from reactive to proactive conservation in the context of accelerating loss of biodiversity.

REFERENCES

- [1]. International Union for Conservation of Nature (IUCN), "The IUCN Red List of Threatened Species (Version 2023-1)", IUCN, (2023). https://www.iucnredlist.org
- [2]. A.V. Anzalone, P.B. Randolph, J.R. Davis, et al., "Search-and-replace genome editing without double-strand breaks or donor DNA", Nature, 576(7785), 149–157, (2019). https://doi.org/10.1038/s41586-019-1711-4
- [3]. N.J. Ouborg, C. Pertoldi, V. Loeschcke, R. Bijlsma, P.W. Hedrick, "Conservation genetics in transition to conservation genomics", Trends in Genetics, 26(4), 177–187, (2010). https://doi.org/10.1016/j.tig.2010.01.001
- [4]. K.J. Campbell, et al., "The next generation of rodent eradications: Innovative technologies and tools to improve species specificity and increase their feasibility on islands", Biological Conservation, 185, 47–58, (2015). https://doi.org/10.1016/j.biocon.2014.10.016
- [5]. G.M. Church, "A pragmatic approach to de-extinction", Nature Biotechnology, 31(6), 517–518, (2013). https://doi.org/10.1038/nbt.2591
- [6]. P.A. Cleves, M.E. Strader, L.K. Bay, J.R. Pringle, M.V. Matz, "CRISPR/Cas9-mediated genome editing in a reef-building coral", Proceedings of the National Academy of Sciences, 115(20), 5235–5240, (2018). https://doi.org/10.1073/pnas.1722151115
- [7]. K.M. Esvelt, N.J. Gemmell, "Conservation demands safe gene drive", PLOS Biology, 15(11), e2003850, (2017). https://doi.org/10.1371/journal.pbio.2003850
- [8]. R. Frankham, "Genetics and extinction", Biological Conservation, 126(2), 131–140, (2005). https://doi.org/10.1016/j.biocon.2005.05.002
- [9]. GeneConvene Global Collaborative, "GeneConvene", (2025, July 18). https://www.geneconvenevi.org
- [10]. P.D. Hsu, E.S. Lander, F. Zhang, "Development and applications of CRISPR-Cas9 for genome engineering", Cell, 157(6), 1262–1278, (2014). https://doi.org/10.1016/j.cell.2014.05.010
- [11]. National Academies of Sciences, "Gene drives on the horizon: Advancing science, navigating uncertainty, and aligning research with public values", National Academies Press, (2016). https://doi.org/10.17226/23405
- [12]. A.J. Piaggio, G. Segelbacher, P.J. Seddon, L. Alphey, E.L. Bennett, R.H. Carlson, et al., "Is it time for synthetic biodiversity conservation?", Trends in Ecology & Evolution, 32(2), 97–107, (2017). https://doi.org/10.1016/j.tree.2016.10.016
- [13]. Revive &Restore, "Reviving the Black-Footed Ferret with CRISPR", (2021). https://reviverestore.org/projects/black-footed-ferret/
- [14]. B. Shapiro, "How to clone a mammoth: The science of de-extinction", Princeton University Press, (2015).
- [15]. M.J.H. van Oppen, J.K. Oliver, H.M. Putnam, R.D. Gates, "Building coral reef resilience through assisted evolution", Proceedings of the National Academy of Sciences, 112(8), 2307–2313, (2015). https://doi.org/10.1073/pnas.1422301112
- [16]. K.M. Whitworth, R.R. Rowland, C.L. Ewen, et al., "Gene-edited pigs are protected from porcine reproductive and respiratory syndrome virus", Nature Biotechnology, 34(1), 20–22, (2016). https://doi.org/10.1038/nbt.3434